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Research Paper

Chemopreventive Potential of Resveratrol in Mouse Skin Tumors Through Regulation of Mitochondrial and PI3K/AKT Signaling Pathways

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Purpose. To investigate the chemopreventive potential of resveratrol, a phytoalexin found in seeds and skin of grapes, berries and peanuts in 7,12 dimethyl benz(a)anthracene (DMBA) induced mouse skin tumorigenesis.
Methods. Topical treatment of resveratrol was given to the animals 1 h prior to DMBA for 28 weeks. At the end of the study period, the skin tumors were dissected out and western blotting was carried out to examine the regulation of proteins involved in anti-tumorigenesis in response to resveratrol.
Results. Chemopreventive properties of resveratrol were reflected by delay in onset of tumorigenesis, reduced cumulative number of tumors, and reduction in tumor volume. Results of the western blotting showed that resveratrol treatment increased the DMBA suppressed p53 and Bax while decreased the expression of Bcl-2 and Survivin. Further, resveratrol supplementation resulted in release of cytochrome C, caspases activation, increase in apoptotic protease-activating factor-1 (Apaf-1) as mechanism of apoptosis induction. Resveratrol was also found to inhibit skin tumorigenesis through regulation of Phosphatidylinositol-3-kinase (PI3K)/ and AKT proteins which are implicated in cancer progression because it stimulates proliferation and suppresses apoptosis.
Conclusions. Based on the results we can conclude that resveratrol regulates apoptosis and cell survival in mouse skin tumors as mechanism of chemoprevention hence deserve to be a chemopreventive agent.

KEY WORDS: apoptosis; chemoprevention; mouse skin tumorigenesis; PI3K/AKT pathway; resveratrol.

INTRODUCTION

Interest in the concept and practice of chemoprevention as an approach to the control of cancer has increased in the recent past. Chemoprevention by naturally occurring agents is gaining much attention as a newer dimension in the management of cancer (1). Many naturally occurring agents have shown cancer chemopreventive potential in a variety of bioassay systems and animal models, having relevance to human disease (2). Resveratrol chemically known as 3, 4′, 5-trihydroxystilbene is a polyphenolic antioxidant compound and is present in red wine, grapes, berries, peanuts etc. Resveratrol is currently under investigation in clinical trials to test the safety and efficiency of resveratrol in the treatment of early stages of cancer. Studies have revealed that resveratrol possess chemopreventive activity against all the three major stages of carcinogenesis i.e. initiation, promotion and progression (3–5). It is reported that resveratrol acts as an antioxidant and antimutagen, induces phase II drug metabolizing enzymes, mediates anti-inflammatory effects, inhibits cyclooxygenase and hydroperoxide functions and induces human promyelocytic leukemia cell differentiation (6–7). The last decade has seen an extraordinary increase in our understanding of apoptosis, and its contribution to cancer and cancer therapy. Several studies have shown that the cancer chemopreventive activity of naturally occurring agents could be attributed to its ability to trigger apoptosis (8–10).

Phosphatidylinositol-3-kinase (PI3K) also plays a pivotal role in several cell signaling networks, including cell cycle progression, differentiation, survival, invasion and metastasis. Several biological effects of PI3K are mediated through activation of its downstream target AKT. AKT is emerging as a central player in tumorigenesis. The serine/threonine kinase AKT is implicated in cancer progression because it stimulates proliferation and suppresses apoptosis (11).

Earlier, we showed that resveratrol induces apoptosis through mitochondrial pathway in mouse skin tumorigenesis taking DMBA as an initiator (12). In this study, we have investigated the chemopreventive potential of resveratrol in mouse skin tumorigenesis taking DMBA as a complete carcinogen. Apart from targeting mitochondrial pathway of apoptosis as mechanism of cancer chemoprevention we have also targeted the proteins involved in PI3K/AKT pathway as it has a strong implication in cancer progression. Here, the study was designed to notably uncover whether resveratrol can promote apoptosis either/both by directly triggering apoptosis-promoting signaling cascades and by blocking signal transduction through the PI3K/AKT as mechanism of cancer chemoprevention.

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MATERIALS AND METHODS

Materials

DMBA, resveratrol, and β-actin were purchased from Sigma Chemical Company (St. Louis, CA, USA). Bcl-2 (ab-2) rabbit polyclonal IgG and Bax (ab-1) rabbit polyclonal IgG, antibodies were procured from Oncogene Research Products (Cambridge, USA). Survivin, caspase 3, caspase 9, cytochrome C, phospho-Akt (Ser473), Poly (ADP-ribose) polymerase (PARP), p85-PI3K, Apaf1, p53 antibodies were procured from Cell Signaling Technology (Beverly, MA, USA). The rabbit anti mouse horseradish peroxidase or goat anti rabbit horseradish peroxidase conjugate secondary antibodies were obtained from Bangalore Genei (Bangalore, India). The polyvinylidene fluoride (PVDF) membranes were obtained from Millipore (Billerica, MA, USA). The rest of the chemicals were of analytical grade of purity and were procured locally.

Animal Bioassay

In order to determine the strength or chemopreventive activity of resveratrol, small laboratory rodents were employed. Female, Swiss albino mice (*Mus musculus L.*; 10–12 gm body weight) were obtained from the Indian Institute of Toxicology Research (Lucknow, India) animal breeding colony. The ethical approval for the experiment was obtained from institutional ethical committee. The animals were caged in polypropylene cages and housed 20 animals per cage on wood chip bedding in an air-conditioned (temperature 23± 2°C, relative humidity 55±5%) animal room. Animals were quarantined for 1 week on a 12/12 h light/dark cycle and were fed solid pellet diet (Crude protein 24%, ether extract 4%, crude fibre 4%, calcium 1%, phosphorous 0.6%, ash 8%, nitrogen source 50%; Ashirwad,Chandigarh, India) and water *ad libitum*. The mouse skin tumors were obtained by using DMBA as a complete carcinogen as described earlier (13). In brief, DMBA/resveratrol was applied topically on shaved dorsal skin in the interscapular region of 2 cm^2. The animals were divided into five groups comprising 20 animals each. For treatment, animals of group I (vehicle control) were only applied with acetone (200 µl) topically. Animals of group II were applied DMBA (5 µg/animal) in acetone (200 µl) topically, which served as a positive control. In order to study the chemopreventive effects of different doses of resveratrol, animals of group III and group IV were topically applied resveratrol 25 µM/animal and 50 µM/animal respectively in acetone (200 µl) 1 h prior to DMBA. Group V animals were only applied resveratrol (50 µM/animal) topically in acetone (200 µl). Treatment was given thrice a week for 28 weeks.

Animals from all the groups were examined throughout the experiment for gross morphological changes locally on skin, including loss of fur and development of tumors. Average tumor volume was calculated using the formula $V = D \times d^2\ \pi/6$, where '*D*' is the biggest dimension of the tumor and '*d*' is the smallest dimension of the tumor. Cumulative number of tumors were also counted to find out the effect of different doses of resveratrol on tumor development. After completion of the study period (28 weeks), all the animals were sacrificed 24 h after the last treatment. Skin from the painted area (with or without tumors) was excised, cleaned, and snap frozen in liquid nitrogen, and stored at −80°C until further use for western blotting.

Preparation of Tissue Lysate

The skin of untreated animals and tumor tissue of tumor bearing animals was removed with sharp scalpel blades, and fat was scrapped off, on ice. The samples were then homogenized in ice-cold lysis buffer (50 mM Tris–HCl, 150 nM NaCl, 1 mM EGTA, 1 mM EDTA, 20 mM NaF, 100 mM Na_3VO_4, 0.5% NP-40, 1% Triton X-100, 1 mM PMSF, 10 µg/ml aprotinin, 10 µg/ml leupeptin, pH 7.4), which were then placed over ice for 30 min (14). The lysate was collected in a microfuge tube and passed through a 21G needle to break up the cell aggregates. The lysates was cleared by centrifugation at 14,000×*g* for 15 min at 4°C and the supernatant (total tissue lysate) was either used immediately or stored at −80°C.

Isolation of Mitochondrial and Cytosolic Fractions

For the determination of release of cytochrome C, the cytosolic and mitochondrial fractions were isolated from uninvolved skin and tumor tissues according to the protocols described by Johnson and Lardy (15). Briefly, tissues were homogenized in buffer containing 0.25 M sucrose and 1 mM EDTA (pH 7.4). The homogenate was centrifuged at 900×*g* for 10 min. The supernatant was centrifuged at 10,000×*g* for 15 min to pellet the mitochondria. The supernatant was further centrifuged at 100,000×*g* to remove any other particulate material (microsomal fraction). The resulting supernatant was designated as the cytosol.

Western Blotting

Western blotting was carried out as described earlier (16). Protein concentration was estimated by the method of Lowry *et al.* (17) using BSA as a standard. Proteins (100 µg) were resolved on 10% sodium dodecyl sulphate (SDS)-polyacrylamide gels and electroblotted on PVDF membranes. The blots were blocked overnight with 5% nonfat dry milk and probed with various monoclonal and polyclonal antibodies at dilutions recommended by the suppliers. Immunoblots were detected through chemiluminescence using enhanced chemiluminescence reagents obtained from Millipore (Billerica, MA, USA). To quantify equal loading, membranes were reprobed with β-actin antibody. Data is presented as the relative density of protein bands normalized to β-actin. The intensities of the bands were quantitated using UN-SCAN IT software (Orem, UT, USA).

Statistical Analysis

For the statistical analysis of skin tumor appearance dynamics, the Kaplan–Meir method of tumor free survival estimation was applied. One-way ANOVA was used between different treated groups after ascertaining the homogeneity of variance between treatments. Post hoc analysis for comparing the two groups was done using the least statistical difference (LSD) technique.

RESULTS

The Chemopreventive Effect of Resveratrol on DMBA Induced Mouse Skin

The results showed the chemopreventive activity of resveratrol on DMBA induced mouse skin tumorigenesis. The animal bioassay revealed a significant (at least 25% difference was considered as significant) delay in the onset of tumorigenesis in resveratrol supplemented groups as compared to the group exposed to DMBA alone. The induction of first tumor was observed on 52nd day in DMBA exposed animals (group II) but the onset of tumorigenesis was observed on 73rd and 79th day in resveratrol supplemented group III and group IV respectively, showing its dose dependency (Table I). The chemopreventive potential of resveratrol was also evident by increase in tumor free survival of animals. Results showed that 100% tumorigenicity was not achieved in resveratrol supplemented animals (group III and IV) till the end of experimental period i.e. 28 weeks. A significant percent of tumor free survival of animals were observed by resveratrol treatment. About 35% of animals remained tumor free in low dose resveratrol supplemented group (group III) while 45% animals remained tumor free in high dose resveratrol supplemented group (group IV; Fig. 1a). Protection could also be seen in terms of reduction in tumor volume. The tumor volume was 98±10 mm^3 tumor volume/mouse in DMBA group, but it was only 48±5 and 34±4 mm^3 in resveratrol supplemented group III and group IV respectively (Fig. 2). Thus, resveratrol supplementation resulted in 51% (group III) and 65% (group IV) suppression in tumors volume. The chemopreventive effect of resveratrol was also evident in terms of reduction in the cumulative number of tumors (CNT) and average number of tumor per tumor bearing mouse. The CNT in group II was 194 at the time of the termination of experiment. The CNT was 73 and 40 in groups III and IV, respectively (Fig. 1b) Thus, resveratrol treatment resulted in 62% (group III) and 79% (group IV) reduction in tumors induced by DMBA (Table I). Similarly, in terms of average number of tumors per tumor bearing mouse, topical treatment of resveratrol resulted in 5.6±1.63 and 3.6±0.46 tumors/tumor bearing mouse in group III and group IV respectively in comparison to DMBA treated group with 9.7±2.6 tumors/tumor bearing mouse (Table I). Data indicates a significant ($p<0.05$) decrease in average number of tumors per tumor bearing mouse in resveratrol treated animals (group III and IV) compared to DMBA treated group.

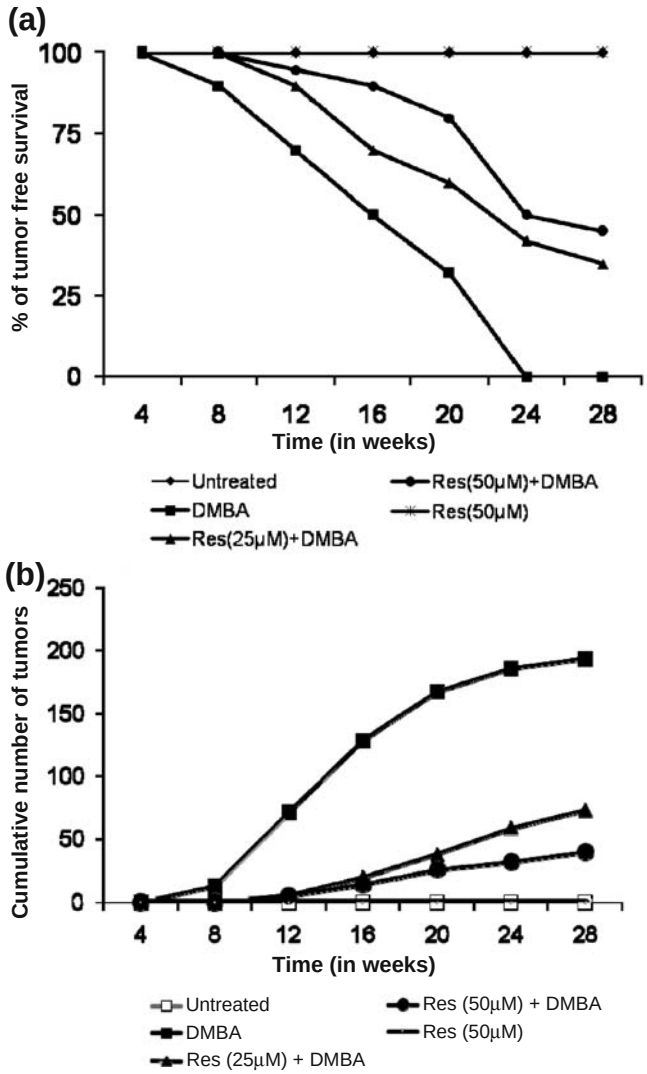

Fig. 1. **A** Kaplan–Meir curve for the determination of tumor free survival by resveratrol treatment on DMBA induced tumorigenesis. The vertical axis shows the percentage of tumor free survival and the horizontal axis shows the weeks of treatment. **B** Effect of resveratrol on incidence of tumorigenesis in terms of cumulative number of tumors.

Table I. Effect of Resveratrol on DMBA Induced Mouse Skin Tumorigenesis

Groups	Treatment*	Ist Induction of tumor (in days)	Number of Animals with tumors	% of animals with tumors	Total tumors (CNT)	Avg. tumor/ tumor bearing mouse (Mean ± SE)
I	Acetone	–	0/20	0	–	–
II	DMBA	52	20/20	100**	194**	9.7±2.6**
III	Resveratrol (25 μM)+DMBA	73	13/20	65*	73*	5.6±1.63*
IV	Resveratrol (50 μM)+DMBA	79	11/20	55*	40*	3.6±0.46*
V	Resveratrol (50 μM)	–	0/20	**0**	**–**	–

Details of treatment are provided in "MATERIALS AND METHODS" section.
*$p<0.05$, significant decrease over DMBA treated group II; **$p<0.05$, significant increase over control group I

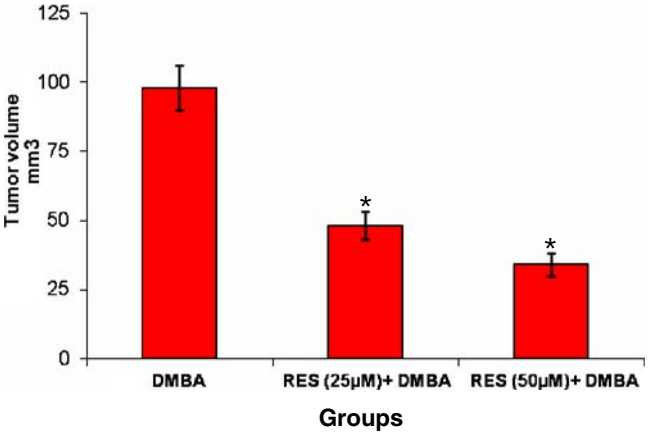

Fig. 2. Effect of resveratrol on tumor volume induced by DMBA. The vertical axis shows the average tumor volume and the horizontal axis shows the treatment. *Asterisk* Values represents significant decrease over DMBA treated group ($p<0.05$).

Resveratrol Induced Expression of p53, its Downstream Regulator Bax and Suppressed the Anti-apoptotic Bcl-2 Expression

Western blot analysis revealed that topical application of resveratrol could effectively modulate the expression level of p53 in comparison to DMBA exposed mouse skin. p53 is critical for apoptosis and lack of its expression or function is associated with an increased risk of tumor formation (18,19). We observed a marked decrease in expression of p53 in DMBA exposed mouse skin (group II) over control (group I). However, resveratrol supplementation up regulated the expression of p53 in group III and IV. No significant ($p<0.05$) difference in the expression levels of p53 was observed between the control group I and V (Fig. 3a).

Mitochondrial permeability leads to the release of apoptogenic factors which is promoted by Bax, a downstream regulator of p53 while Bcl-2 inhibits its effect. We further, ascertained the modulating effect of resveratrol on Bcl-2 family proteins. Bax was down regulated in DMBA exposed group II while its expression was enhanced in resveratrol supplemented groups (group III and IV; Fig. 3b). Conversely, Bcl-2 was over-expressed in DMBA treated animals (group II). The DMBA induced expression of Bcl-2 was down regulated in resveratrol supplemented animals (group III and IV; Fig. 3c).

Resveratrol Induces Apoptosis via Cytochrome C Release, Caspase Activation and PARP Cleavage

Mitochondrial permeability leads to the release of apoptogenic factors like cytochrome C, Apaf 1 and activation of caspase 3, caspase 9, and PARP cleavage. Release of cytochrome C into cytosol and expression of Apaf-1 were down regulated in DMBA exposed animals over control (group I). Resveratrol treatment elevated level of cytosolic cytochrome C (Fig. 3d) and expression level of Apaf-1 (Fig. 3e) in comparison to DMBA treated group (group II). However, results of western blotting also showed cleavage of

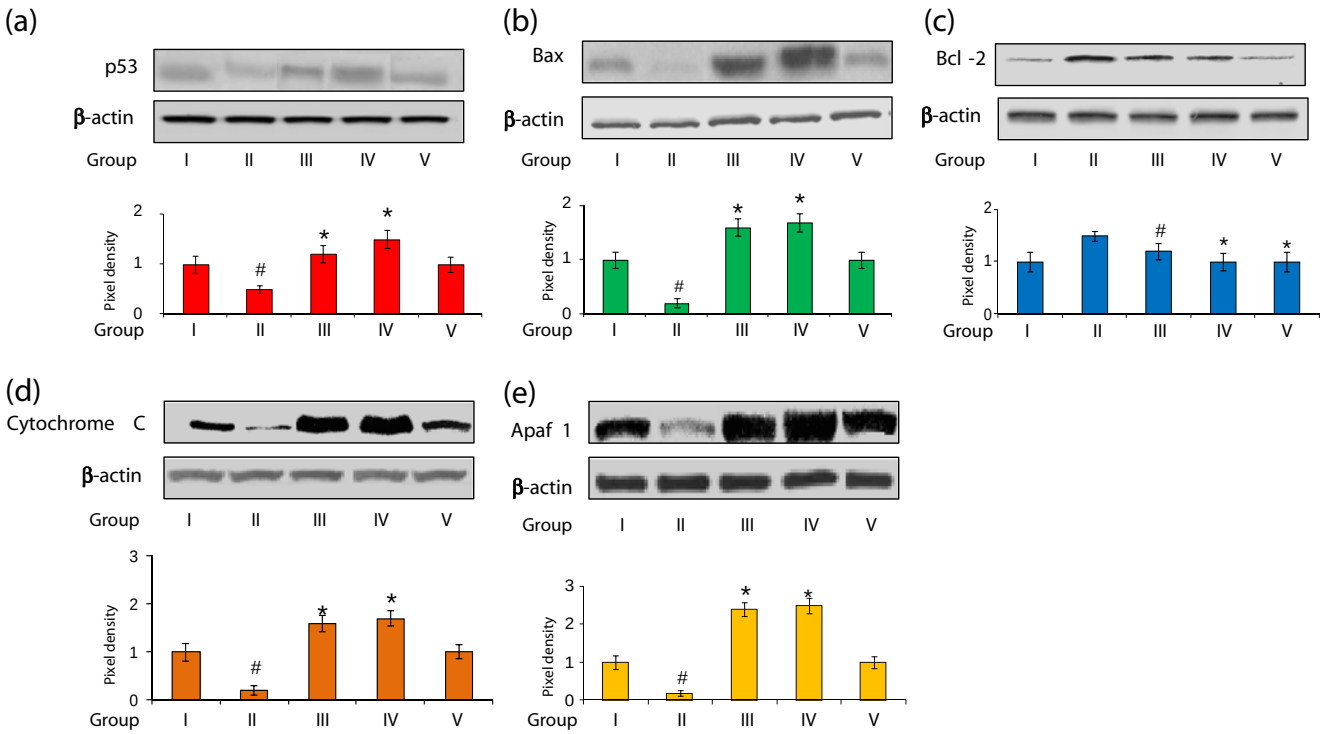

Fig. 3. Western blots showing the effect of resveratrol on **a** p53, **b** Bax, **c** Bcl-2, **d** cytochrome C, **e** Apaf-1 in mouse skin/tumors in different groups. *Group I* Acetone, *Group II* acetone+DMBA, *Group III* resveratrol (25 μM)+DMBA, *Group IV* resveratrol (50 μM)+DMBA, *Group V* acetone+resveratrol (50 μM). Equal loading was confirmed by reprobing the membrane with β-actin. The *bands* shown here are from a representative experiment repeated three times with similar results. *Pound sign* Value is significantly different over group I, $p<0.01$. *Asterisk* Values are significantly different over group II, $p<0.05$. The standard errors of pixel densities of bands are represented by *error bars*.

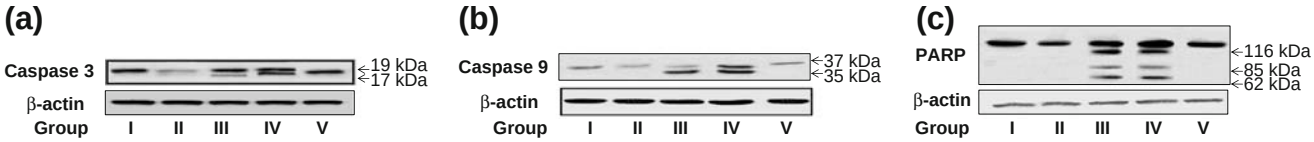

Fig. 4. Western blots showing the effect of resveratrol on **a** caspase 3 **b** caspase 9 **c** PARP in mouse skin/tumors in different groups. *Group I* Acetone, *Group II* acetone+DMBA, *Group III* resveratrol (25 μM)+DMBA, *Group IV* resveratrol (50 μM)+DMBA, *Group V* acetone+resveratrol (50 μM). Equal loading was confirmed by reprobing the membrane with β-actin. The *bands* shown here are from a representative experiment repeated three times with similar results. The standard errors of pixel densities of bands are represented by *error bars*.

caspase-9 (35 and 37 kDa) and caspase-3 (19 and 17 kDa) and PARP (116, 85, and 62 kDa) on resveratrol treatment (Fig. 4).

Resveratrol Induced Down Regulation of Survivin and PI3K/AKT Pathway

We further studied the effect of resveratrol treatment on survivin, a member of inhibitor of apoptosis. The levels of survivin were over-expressed in DMBA exposed group (group II) over controls. A comparatively low level of expression of survivin was recorded in resveratrol supplemented groups (group III and IV; Fig. 5a).

Further, we also studied the effect of resveratrol treatment on p85-PI3K and phospho-AKT (Ser473; protein Kinase B) proteins whose abnormal signaling contributes cancer progression. Their expression was found to be increased in DMBA treatment animals. However, expression of both the proteins was down regulated following resveratrol supplementation (Fig. 5b,c).

DISCUSSION

Cancer chemoprevention by using nontoxic chemical substances is regarded as a promising alternative strategy for control of human cancer. In recent years, many naturally occurring substances have been shown to protect against experimental carcinogenesis (20,21) In this regard, resveratrol (3,5,4′-trihydroxy stilibene), a phytoalexin found in a multitude of dietary plants including grapes and peanuts have been shown to provide cancer chemopreventive effects in both *in vivo* and *in vitro* systems (7). One of their main properties of resveratrol is their antioxidant activity, which enables them to attenuate the development of atherosclerosis, inflammatory diseases, and cancer (22).

In the present investigation, the chemopreventive activity of resveratrol was studied by employing mouse skin carcinogenesis model. The animal bioassay revealed a significant delay in onset of tumorigenesis, significantly reduced cumulative number of tumors, and significant reduction in tumor volume (at least 25% difference was considered as significant). Thus, overall tumorigenesis experiment clearly showed a strong protective effect of resveratrol against DMBA induced mouse skin tumorigenesis. This chemopreventive potential of resveratrol may be associated with inhibition of mutation. It has been shown that resveratrol inhibited methylmethansulfonate and benzopyrene induced reversion *Salmonella typhimurium* TA100 and also prevented cyclophosphamide induced micronucleus formation of mice bone marrow (23) indicating its antimutagenicity.

We further extended this work to gain insight into the signaling network and interaction points modulated by resveratrol via ascertaining their role in modulation of the proteins involved in the mitochondrial pathway of apoptosis and protein kinase B pathway. The prevention of cancer is profoundly dependent on the p53 tumor suppressor protein. The ability of p53 to eliminate excess, damaged or infected cells by apoptosis is vital for the proper regulation of cell proliferation in multi-cellular organisms (24). p53 is activated by external and internal stress signals that promote its nuclear accumulation in an active form. In turn, p53 induces either viable cell growth arrest or apoptosis. The latter activity is crucial for tumor suppression.

p53 participates in apoptosis induction by acting directly at mitochondria. Localization of p53 to the mitochondria occurs in response to apoptotic signals and precedes cytochrome C release and caspase-3 activation. Recently, Mihara *et al.* (25) also extended this finding to show that p53 promotes permeabilization of the outer mitochondrial membrane by forming complexes with the protective Bcl-XL and

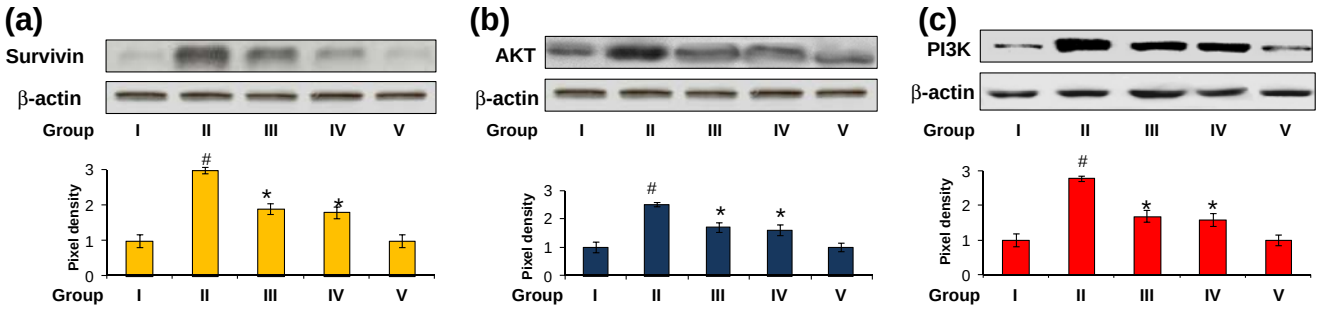

Fig. 5. Western blots showing the effect of resveratrol on **a** Survivin **b** AKT **c** PI3K in mouse skin/tumors in different groups. *Group I* Acetone, *Group II* acetone+DMBA, *Group III* resveratrol (25 μM)+DMBA, *Group IV* resveratrol (50 μM)+DMBA, *Group V* acetone+resveratrol (50 μM). Equal loading was confirmed by reprobing the membrane with β-actin. The *bands* shown here are from a representative experiment repeated three times with similar results. *Pound sign* Value is significantly different over group I, $p<0.01$.*Asterisk* Values are significantly

Bcl-2 proteins. Transcriptional activation of mitochondrial proteins, such as Bax causes apoptosis and p53 down regulates Bcl-2 which acts as an anti-apoptogenic agent (26). In our study, we found that the expression of p53 and its downstream regulator Bax was increased and Bcl-2 was decreased on resveratrol supplementation. Increased expression of Bax can induce apoptosis by suppressing the activity of Bcl-2 (27,28), confirming Bcl-2 and Bax is crucial for the apoptosis induced by chemopreventive agents (29). Interaction among the Bcl-2 family proteins (Bax, Bak, Bcl-2, Bcl-X, etc) stimulates the release of cytochrome C which promotes the formation of apoptosome with Apaf1 which in turn activates executioner caspases to orchestrate apoptosis. Caspases are the crucial components of the apoptosis pathway. The important step in activation of the cell death program is the activation of caspase 3 and caspase 9 (30). PARP is a protein involved in a number of cellular processes involving mainly DNA repair and programmed cell death (31). Consistent with the above studies, our study also showed the up-regulation of the proteins like cytochrome C, Apaf1, caspase 9, caspase 3 and PARP in resveratrol supplemented groups in comparison to DMBA treated group. The ability of PARP is to repair damaged DNA which is prevented through its cleavage by executioner caspases (32). Survivin proteins, member of inhibitor of apoptosis, directly inhibits apoptosis and its expression is found to frequently high in cancer cells and correlated with resistance to chemotherapy (33,34). Survivin has a role in preventing apoptosis, possibly by impairing caspases activation and mitochondrial dysfunction (35). Similar observation was found in our study, showing increased level of Survivin in DMBA exposed group. However, resveratrol treatment modulated the survivin protein and inducing apoptosis probably through activation of caspases. Also, anticancer potential of resveratrol was observed by growth inhibition mediated through up regulation of p53 and Bax; down-regulation of Bcl-2 and Survivin and activation of caspases in breast, pancreas, skin and prostate cancer (12,36).

PI3K is a lipid kinase which plays a central role in signaling pathways important to biological processes including cell survival, proliferation, cell growth and cell motility (37). PI3K generates phosphatidylinositol-3, 4, 5-trisphosphate, a second messenger essential for the translocation of AKT to the plasma membrane where it is phosphorylated at serine 473 and activated by phosphoinositide-dependent kinase PDK 1 and PDK2. Activation of AKT by phosphorylation at serine 473 plays a pivotal role in fundamental cellular functions such as cell proliferation and cell survival by phosphorylating a variety of substrates. In recent years, it has been reported that altered PI3K/AKT signaling pathway frequently occur in human cancer. Several studies have reported the involvement of PI3K/AKT signaling pathway in resveratrol induced growth inhibition of different cell types (38–40).

In our study, decreased expression of regulatory p85-PI3K and associated proteins phospho-AKT by resveratrol treatment support the inhibition in development of tumors. Several small molecules designed to specifically target PI3K/AKT have been developed and induced cell cycle arrest or apoptosis in human cancer cells *in vitro* and *in vivo* (41). Therefore, specific inhibition of the activation of AKT by phosphorylation at serine 473 may be a valid approach for treatment of human malignancies. The expression of regulatory p38-PI3K protein and phospho-AKT (Ser473) was found to be decreased in resveratrol supplemented groups over DMBA exposed group. Thus, our study convincingly shows role of resveratrol in modulation of the proteins involved in the protein kinase B pathway.

Because the activation of the PI3K/Akt pathway leads to increased expression of Bcl-2 (42), we assessed the effect of resveratrol on Bcl-2 family proteins, and our data showed that resveratrol treatment (a) down-regulates Bcl-2 protein, and (b) up-regulates Bax protein. Furthermore, our data also showed a release of cytochrome C in resveratrol-treated groups suggesting that inhibition of Akt activation is possibly preceded by modulations in mitochondrial damage leading to a shift in the balance between proapoptotic and antiapoptotic proteins in favor of apoptosis. The cancer chemopreventive potential of resveratrol may be mediated through cell cycle arrest which further leads to apoptosis. Adhami *et al.* (21) showed that resveratrol causes a downregulation of hyperphosphorylated retinoblastoma protein with a relative increase in hypophosphorylated retinoblastoma that, in turn, compromises with the availability of free transcription factor E2F those regulate the progression of the cell cycle and finally apoptosis.

CONCLUSIONS

Altogether the results of the present investigations showed chemopreventive effects of resveratrol through the regulation of the expression of proteins involved in mitochondrial pathway of apoptosis and protein kinase B pathway in DMBA induced mouse skin carcinogenesis. So from this study we can conclude that resveratrol can regulate apoptotic pathways both by directly triggering apoptosis-promoting signaling cascades and by blocking signal transduction through the PI3K/AKT as mechanism of cancer chemoprevntion. It is hopeful that further characterization of pathways regulating cell cycle progression and apoptosis will facilitate novel drug discovery programs to exploit resveratrol for the prevention and treatment of several human cancers.

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REFERENCES

1. D. R. Bickers, and M. Athar. Novel approaches to chemoprevention of skin cancer. *J. Dermatol.* **27**:691–695 (2000).
2. L. Fremont. Biological effects of resveratrol. *Life Sci.* **66**:663–673 (2000). doi:10.1016/S0024-3205(99)00410-5.
3. D. K. Das, M. Sato, P. S. Ray, G. Maulik, R. M. Engleman, A. A. Bertelli, and A. Bertelli. Cardio protection of red wine: role of polyphenolic antioxidants. *Drugs Exp. Clin. Res.* **25**:115–120 (1999).
4. U. R. Pendurthi, F. Meng, N. Mackman, and L.V. Rao. Mechanism of resveratrol mediated suppression of tissue factor gene expression. *Thromb. Haemost.* **87**:155–162 (2002).

5. S. Banerjee, C. Bueso-Ramos, and B.B. Aggarwal. Suppression of 7, 12-dimethylbenz(a)anthracene-induced mammary carcinogenesis in rats by resveratrol: role of nuclear factor—Kappa B, cyclooxygenase 2, and matrix metalloprotease 9. *Cancer Res.* **62**:4945–4954 (2002).
6. M. Jang, L. Cai, O. G. Udeani, K. V. Slowing, C. F. Thomas, C. W. W. Beecher, H. H. S. Fong, N. R. Farnsworth, A. D. Kinghorn, R. G. Mehta, R. C. Moon, and J. M. Pezzuto. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science.* **275**:218–220 (1997). doi:10.1126/science.275.5297.218.
7. M. V. Clement, J. L. Hirpara, and S. Pervaiz. Chemopreventive agent resveratrol, a natural product derived from grapes triggers Cd95 signalling-dependent apoptosis in human tumor cells. *Blood.* **92**:996–1002 (1998).
8. Y. J. Surh, Y. J. Hurh, J. Y. Kang, E. Lee, G. Kong, and S. J. Lee. Resveratrol,an antioxidant present in red wine, induces apoptosis in human promyelocytic leukemia (HL-60) cells. *Cancer Lett.* **140**:1–10 (1999). doi:10.1016/S0304-3835(99)00039-7.
9. Q. B. She, A. M. Bode, W. Y. Ma, N. Y. Chen, and Z. Dong. Resveratrol-induced activation of p53 and apoptosis is mediated by extracellular-signal-regulated protein kinases and p38 kinase. *Cancer Res.* **61**:1604–1610 (2001).
10. N. Ahmad, V. M. Adhami, F. Afaq, D. K. Feyes, and H. Mukhtar. Reveratrol causes WAF-1/p21-mediated G1-phase arrest of cell cycle and induction of apoptosis in human epidermoid carcinoma A431 cells. *Clin. Cancer Res.* **7**:1466–1473 (2001).
11. R. Joseph, and B. Alfanso. AKT plays a central role in tumorigenesis. *Proc. Natl. Acad. Sci.* **98**:10983–10985 (2001). doi:10.1073/pnas.211430998.
12. N. Kalra, P. Roy, S. Prasad, and Y. Shukla. Resveratrol induces apoptosis involving mitochondrial pathways in mouse skin tumorigenesis. *Life Sci.* **82**:348–358 (2008). doi:10.1016/j.lfs.2007.11.006.
13. A. Singh, and Y. Shukla. Anti tumor activity of diallyl sulphide on polycyclic aromatic hydrocarbon-induced mouse skin carcinogenesis. *Cancer Lett.* **131**:209–214 (1998). doi:10.1016/S0304-3835(98)00152-9.
14. I. A. Siddiqui, V. M. Adhami, F. Afaq, N. Ahmad, and H. Mukhtar. Modulation of phosphatidylinositol-3-kinase/protein kinase B- and mitogen-activated protein kinase-pathways by tea polyphenols in human prostate cancer cells. *J. Cell Biochem.* **91**:232–242 (2004). doi:10.1002/jcb.10737.
15. D. Johnson, and H. Lardy. In R. W. Estabrook, and M. E. Pullman (eds.), Methods in Enzymology, Oxidation and Phosphorylation, Vol. X, Academic, New York, 1967, pp. 94–96.
16. A. Arora, I. A. Siddiqui, and Y. Shukla. Modulation of p53 in 7,12-dimethylbenz[a] benzanthracene-induced skin tumors by diallyl sulphide in Swiss albino mice. *Mol. Cancer Ther.* **11**:1459–1466 (2004).
17. O. H. Lowry, N. K. Rosenbrough, and A. L. Farr. Protein measurement with folin phenol reagent. *J. Biol. Chem.* **193**:265–275 (1951).
18. S. W. Lowe, H. E. Ruley, T. Jacks, and D. E. Housman. p53-dependent apoptosis modulates the cytotoxicity of anticancer drugs. *Cell.* **74**:957–967 (1993). doi:10.1016/0092-8674(93)90719-7.
19. A. R. Clarke, S. Gledhill, M. L. Hooper, C. C. Bird, and A. H. Wyllie. p53 dependence of early apoptotic and proliferative responses within the mouse intestinal epithelium following gamma-irradiation. *Oncogene.* **9**:1767–1773 (1994).
20. G. J. Kapadia, M. A. Azuine, H. Tokuda, M. Takasaki, T. Mukainaka, T. Konoshima, and H. Nishino. Chemopreventive effectof resveratrol, sesamol, sesame oil and sunflower oil in the Epstein-barr virus early antigen activation assay and the mouse skin two stage carcinogenesis. *Pharmacol. Res.* **45**:499–505 (2002). doi:10.1006/phrs.2002.0992.
21. V. M. Adhami, F. Afaq, and N. Ahmad. Involvement of the Retinoblastoma (pRb)-E2F/DP pathway during antiproliferative effects of resveratrol in human epidermoid carcinoma (A431) Cells. *Biochem. Biophys. Res. Commun.* **288**:579–585 (2001). doi:10.1006/bbrc.2001.5819.
22. G. J. Soleas, L. Grass, P. D. Josephy, D. M. Goldberg, and E. P. Diamandis. A comparison of the anticarcinogenic properties of four red wine polyphenols. *Clin. Biochem.* **35**:119–124 (2002). doi:10.1016/S0009-9120(02)00275-8.
23. Z. D. Fu, Y. Cao, K. F. Wang, S. F. Xu, and R. Han. Chemopreventive effect of resveratrol to cancer. *Ai Zheng.* **23**:869–873 (2004).
24. J. F Kerr, A. H. Wyllie, and A. R. Currie. Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br. J. Cancer.* **26**:239–257 (1972).
25. M. Mihara, S. Erster, A. Zaika, O. Petrenko, T. Chittenden, P. Pancake, and U. M. Moll. p53 has a direct apoptogenic role at the mitochondria. *Mol. Cell.* **11**:77–590 (2003). doi:10.1016/S1097-2765(03)00050-9.
26. H. J. Harn, L. I. Ho, C. A. Liu, and G. C. Liu. Down regulation of bcl-2 by p53 in nasopharyngeal carcinoma and lack of detection of its specific t (14;18) chromosomal translocation in fixed tissues. *Histopathology.* **28**:317–323 (1996). doi:10.1046/j.1365-2559.1996.d01-431.x.
27. D. M. Hockenbery, C. D. Giedt, J. W. O'Neill, and M. K. Manion. Mitochondria and apoptosis: new therapeutic targets. *Adv. Cancer Res.* **85**:203–242 (2002). doi:10.1016/S0065-230X(02)85007-2.
28. L. MacCarthy-Morrogh, A. Mouzakiti, P. Townsend, and M. Brimmell. Bcl-2-related proteins and cancer. *Biochem. Soc. Trans.* **27**:785–789 (1999).
29. G. S. Salomons, H. J. Brady, M. Verwijs-Janssen, and J. D. Van Den Berg. The Bax:Bcl-2 ratio modulates the response to dexamethasone in leukaemic cells and is highly variable in childhood acute leukaemia. *Int. J. Cancer.* **71**:959–965 (1997). doi:10.1002/(SICI)1097-0215(19970611)71:6<959::AID-IJC9>3.0.CO;2-X.
30. G. M. Cohen. Caspases: the executioners of apoptosis. *Biochem. J.* **326**:1–16 (1997).
31. S. Bursztajn, J. J. Feng, S. A. Berman, and A. R. Nanda. Poly (ADP-ribose) polymerase induction is an early signal of apoptosis in human neuroblastoma. *Brain Res. Mol. Brain Res.* **76**:363–376 (2000). doi:10.1016/S0169-328X(00)00026-7.
32. U. Kolthur-Seetharam, F. Dantzer, M. W. McBurney, G. de Murcia, and P. Sassone-Corsi. Control of AIF mediated cell death by the functional interplay of SIRT1 and PARP-1 in response to DNA damage. *Cell cycle.* **5**:873–877 (2006).
33. D. C. Altieri. Survivin, versatile modulation of cell division and apoptosis in cancer. *Oncogene.* **22**:8581–8589 (2003). doi:10.1038/sj.onc.1207113.
34. Q. L. Deveraux, and J. C. Reed. IAP family proteins—suppressors of apoptosis. *Genes Dev.* **13**:239–252 (1999). doi:10.1101/gad.13.3.239.
35. M. Castedo, J. L. Perfettini, T. Roumier, K. Andreau, R. Medema, and G. Kroemer. Cell death by mitotic catastrophe: a molecular definition. *Oncogene.* **23**:2825–2837 (2004). doi:10.1038/sj.onc.1207528.
36. B. B. Aggarwal, A. Bhardwaj, R. S. Aggarwal, N. P. Seeram, S. Shishodia, and Y. Takada. Role of resveratrol in prevention and therapy of cancer: preclinical and clinical trials. *Anticancer Res.* **24**:2783–2840 (2004).
37. T. O. Chan, S. E. Rittenhouse, and P. N. Tsichlis. AKT/PKB and other D3 phosphoinositide-regulated kinases: kinase activation by phosphoinositide-dependent phosphorylation. *Annu. Rev. Biochem.* **68**:965–1041 (1999). doi:10.1146/annurev.biochem.68.1.965.
38. R. Srivastava, A. Ratheesh, R. K. Gude, K. V. Rao, D. Panda, and G. Subrahmanyam. Resveratrol inhibits type II phosphatidylinositol turnover. *Biochem. Pharmacol.* **70**:1048–1045 (2005). doi:10.1016/j.bcp.2005.07.003.
39. T. M. Poolman, L. L. Ng, P. B. Farmer, and M. M. Manson. Inhibition of the respiratory burst by resveratrol in human monocytes: correlation with inhibition of PI3K signalling. *Free Radic. Biol. Med.* **39**:118–132 (2005). doi:10.1016/j.freeradbiomed.2005.02.036.
40. O. Rachid, and M. Alkhalaf. Resveratrol regulation of PI3K-AKT signaling pathway genes in MDA-MB-231 breast cancer cells. *Cancer Genomics Proteomic.* **3**:383–388 (2006).
41. Y. Lu, H. Wang, and G. B. Mills. Targeting PI3K-AKT pathway for cancer therapy. *Rev. Clin. Exp. Hematol.* **7**:205–228 (2003).
42. L. Asnaghi, A. Calastretti, A. Bevilacqua, I. D'Agnano, G. Gatti, G. Canti, D. Delia, S. Capaccioli, and A. Nicolin. Bcl-2 phosphorylation and apoptosis activated by damaged microtubules require mTOR and are regulated by Akt. *Oncogene.* **23**:5781–5791 (2004). doi:10.1038/sj.onc.1207698.